



1,4-BENZODIAZEPIN-2-ONE DERIVED NEUROKININ-1 RECEPTOR ANTAGONISTS.

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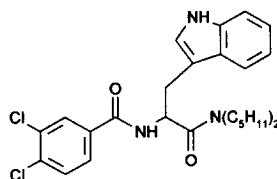
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Abstract The identification of a series of 1,4-benzodiazepin-2-one derived NK₁ receptor antagonists is described and a bioactive conformation is proposed. © 1997 Elsevier Science Ltd.

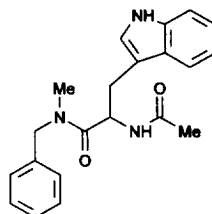
The structural similarities between G-protein coupled 7-transmembrane receptors have been the focus of a great deal of research. The advent of site-directed mutagenesis, used in tandem with photoaffinity labelling studies, has begun to provide the means of studying the key interactions between receptors and their ligands.¹ Intriguingly, the structural similarities between receptors appears to have a corollary with the structural similarities between non-peptide agonists/antagonists that have been discovered for these systems.^{1b} This has led to the concept of so-called "privileged structures" which occur repeatedly in pharmacologically active compounds.² Hence, these key motifs, such as diphenylmethane and 1,4-benzodiazepine, are increasingly being incorporated into combinatorial libraries directed at medicinal targets.³

We have been fascinated for some time by the structural similarities between the gastrin/cholecystokinin (CCK) and neurokinin (NK) receptors and their ligands. Both families have peptide agonists as their natural ligands and both, according to initial evidence, bind non-peptide ligands in a common pocket between the transmembrane helices, at a site distinct from the natural agonist binding site.^{1b} Furthermore, at the simplest level the peptidic antagonists reported for these systems are also highly similar e.g. (1) and (2).⁴



CCK_A IC₅₀ 190nM^{4a}

(1)



NK₁ IC₅₀ 750nM^{4b}

(2)

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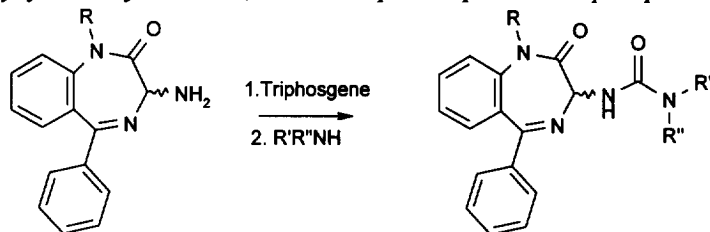
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The classical non-peptide antagonists for the CCK receptors were developed by Freidinger *et al.* by simplifying and refining the structure of the natural product asperlicin to give the 1,4-benzodiazepines L-364,178 and L-365,260, selective antagonists of the CCK_A and CCK_B receptor subtypes respectively.⁵ The benzodiazepine template has subsequently been used by many workers in other fields.⁶ Herein we report the identification of benzodiazepine-derived NK₁ receptor antagonists, NMR studies on these compounds and initial structure-activity relationships around this novel series.

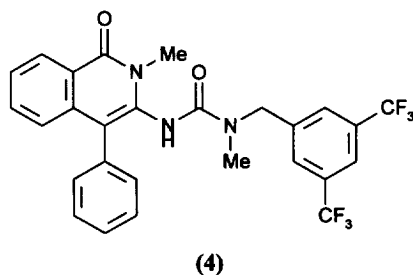
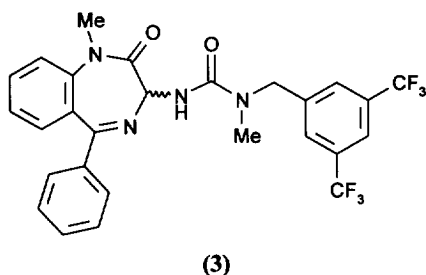
Results and Discussion.

At the outset of our programme the precise structural requirements of the NK₁ receptor were unclear. We therefore screened a wide variety of benzodiazepines which had been previously prepared for our gastrin/CCK programmes but failed to identify any active compounds. Similarly a small library of 50 compounds weighted towards aromatic derivatives (Figure 1) did not produce any active leads ($pK_i > 7$).⁷

Figure 1. Array synthesis of 3-ureido-1,4-benzodiazepines as potential NK₁ receptor antagonists



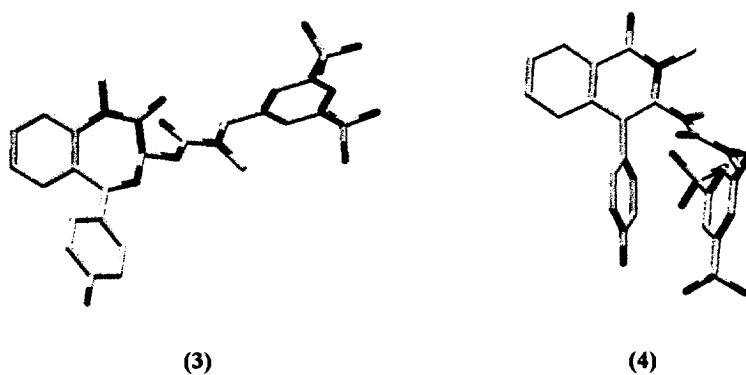
Although libraries of such size and diversity are limited, this set included compound (3) which incorporates the 3,5-bis(trifluoromethyl)benzyl side-chain, a common structural feature in NK₁ receptor antagonists from Merck⁸ and, more recently, in compounds from Takeda such as (4).⁹ The Takeda compound was developed using a strategy which also looked to derive ligands for the NK₁ receptor based on the perceived similarities between the gastrin/CCK and NK₁ receptor antagonists.



The NK₁ receptor has proven to be extremely promiscuous - a wide variety of effective templates have now been described.¹⁰ To date, most of these templates contain two aryl rings which can adopt an orientation that will allow a π - π interaction in at least some of their accessible low-energy conformations;¹¹ for example Takeda have described modelling which shows that an analogue of (4) can lie in just such a conformation (Figure 2).⁹ It has been proposed that this stacking of the two aryl rings is important in the bioactive conformation.¹²

The data available from both X-ray analysis and NMR studies in solution suggest that the C-3 substituted 1,4-benzodiazepines such as (3) tend to adopt an extended conformation^{2a} (Figure 2) keeping the aromatic rings some distance apart, and this may explain the failure to identify a NK₁ receptor antagonist from within this series.

Figure 2. *Low energy conformations of (3) and (4).*



Molecular modelling¹³ of an alternative 1,4-benzodiazepine template (5) suggested that, of the available low energy conformations, (5) has two energetically accessible pseudo-boat conformations in which two of the aromatic rings can adopt an orientation which allows a π - π interaction (Figure 3). Upon chemical synthesis, compound (5) proved to have high affinity for the NK₁ receptor (pK_i 8.0).

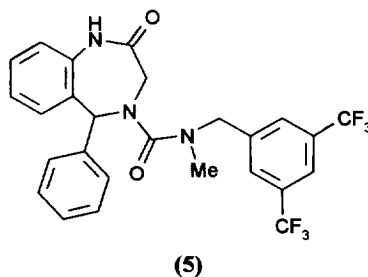
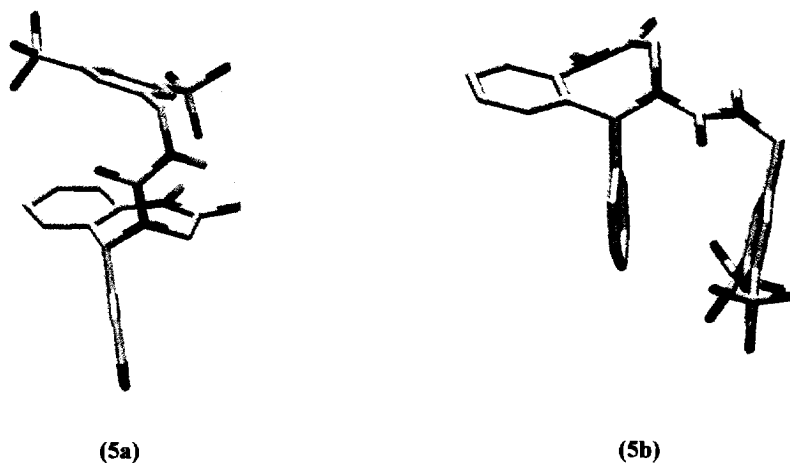
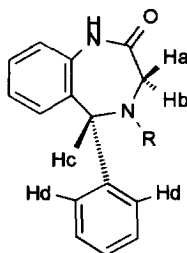


Figure 3. *Low energy conformations of compound (5)*

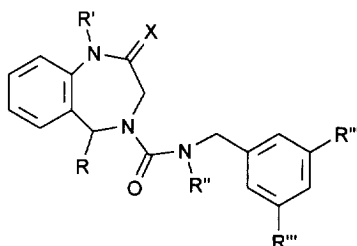
We were keen therefore to identify which of these conformations corresponded to the bioactive conformation. NMR studies on the solution conformation of (5) in CDCl_3 are not consistent with there being a substantial population of conformation 5b - no n.O.e. is observed between protons Ha and Hc. Although the absence of a n.O.e. cannot be considered conclusive, these protons would be in a pseudo 1,3-diaxial relationship in conformation 5b. Furthermore a n.O.e. between Hb and Hd strongly suggests that the compound resides predominantly in conformation 5a. This would suggest that the 5-phenyl group is not essential for high binding affinity. The NMR studies cannot entirely rule out an alternative pseudo-chair conformation, however, modelling and X-ray studies by ourselves and others¹⁴ would seem to make the pseudo-chair less likely.



To confirm this theory we elected to synthesise compound (6) in which the 5-phenyl group is replaced by a 5-methyl group. Compound (6) is only marginally less potent than (5) (pK_i 7.5) and appears to adopt a similar conformation in d_6 -DMSO solution by NMR. This suggests that conformer 5a may be the bioactive conformation.

A range of other simple derivatives were prepared and results are summarised in Table 1. Interestingly the benzodiazepine template is not acting as a passive scaffold - N-methylation of the amide (7) results in a reduction of activity suggesting that the N-1 amide is involved in receptor binding, presumably acting as a hydrogen bond donor. The tetra-substitution of the urea appears to be critical (compare compounds (5) and (9)) possibly by allowing the urea to twist out of a fully planar conformation or by allowing both *cis* and *trans* conformations about the urea.¹⁵ The 3,5-bis(trifluoromethyl) substitution on the benzyl group contributes substantially to potency (compare compounds (5) and (10)).

Table 1 *Structure-activity relationships in the 1,4-benzodiazepin-2-one series of NK₁ receptor antagonists*



Compound No.	R	R'	R''	R'''	X	pK _i ⁷	n	SEM
(5)	Ph	H	Me	CF ₃	O	8.0	3	0.25
(6)	Me	H	Me	CF ₃	O	7.5	4	0.21
(7)	Me	Me	Me	CF ₃	O	6.0	4	0.06
(8)	Me	H	Me	CF ₃	2H	7.4	3	0.04
(9)	Ph	H	H	CF ₃	O	6.2	3	0.05
(10)	Ph	H	Me	H	O	6.0	4	0.06
(11)	CO ₂ Me	H	Me	CF ₃	O	6.5	3	0.24
(12)		H	Me	CF ₃	O	6.1	4	0.26

Conclusion

The 1,4-benzodiazepin-2-one derivative (5) represents the first example of a new class of potent NK₁ receptor antagonists, and provides another example of the use of the 1,4-benzodiazepine template for antagonists of G-protein coupled 7- transmembrane receptors.

References and Notes

1. For example see : a) Gether, U.; Lowe III, J.A.; Schwartz, T.W.; *Biochem. Soc. Trans.*, **1995**, 23, 96. b) Cascieri, M.A.; Fong, T.M.; Strader, C.D.; *Drugs of the Future*, **1996**, 21, 521 and references therein.
2. a) Evans, B.E.; Rittle, K.E.; Bock, M.G.; DiPardo, R.M.; Freidinger, R.M.; Whitter, W.L.; Lundell, G.F.; Veber, D.F.; Anderson, P.S.; Chang, R.S.L.; Lotti, V.J.; Cerino, D.J.; Chen, T.B.; Kling, P.J.; Kunkel, K.A.; Springer, J.P.; Hirshfield, J.; *J. Med. Chem.*, **1988**, 31, 2235. b) Wiley, R.A.; Rich, D.H.; *Med. Res. Rev.*, **1993**, 13, 327.
3. For example: Bunin, B.A.; Plunkett, M.J.; Ellman, J.A.; *Proc. Natl. Acad. Sci.*, **1994**, 91, 4708.
4. a) Kerwin, J.F.; Wagenaar, F.; Kopecka, H.; Lin, C.; Miller, T.; Witte, D.; Stashko, M.; Nadzan, A.M.; *J. Med. Chem.* **1991**, 34, 3350. b) Hipskind, P.A.; Howbert, J.J.; Bruns, R.F.; Cho, S.S.Y.; Crowell, T.A.; Foreman, M.M.; Gehlert, D.R.; Iyengar, S.; Johnson, K.W.; Krushinski, J.H.; Li, D.L.; Lobb, K.L.; Mason, N.R.; Muehl, B.S.; Nixon, J.A.; Phebus, L.A.; Regoli, D.; Simmons, R.M.; Threlkeld, P.G.; Waters, D.C.; Gitter, B.D.; *J. Med. Chem.*, **1996**, 39, 736.
5. For a review of CCK receptor antagonists see: Trivedi, B.K.; *Curr. Med. Chem.*, **1994**, 1, 313.
6. a) Romer, D.; Büscher, H.H.; Hill, R.C.; Maurer, R.; Petcher, T.J.; Zeugner, H.; Benson, W.; Finner, E.; Milkowski, W.; Thies, P.W.; *Nature*, **1982**, 298, 759. b) Papageorgiou, C.; Borer, X.; *Bioorg. Med. Chem. Lett.*, **1996**, 6, 267. c) McDowell, R.S.; Blackburn, B.K.; Gadek, T.R.; McGee, L.R.; Rawson, T.; Reynolds, M.E.; Robarge, K.D.; Somers, T.C.; Thorsett, E.D.; Tischler, M.; Webb, R.R. Venuti, M.C.; *J. Am. Chem. Soc.*, **1994**, 116, 5077.
7. Affinities of compounds for the NK₁ receptor were determined by the method described in Ward P.; Armour D.R.; Bays D.E.; Evans B.; Giblin, G.M.P.; Heron N.; Hubbard T.; Liang K.; Middlemiss D.; Mordaunt, J.; Naylor A.; Pegg N.A.; Vinader, M.V.; Watson S.P.; Bountra, C.; Evans D.C.; *J. Med. Chem.*, **1995**, 38, 4985.
8. a) MacLeod, A.M.; Merchant, K.J.; Brookfield, F.; Kelleher, F.; Stevenson, G.; Owens, A.P.; Swain, C.J.; Cascieri, M.A.; Sadowski, S.; Ber, E.; Strader C.D.; MacIntyre, D.E.; Metzger, J.M.; Ball, R.G.; Baker R.; *J. Med. Chem.*, **1994**, 37, 1269. b) Harrison, T.; Williams, B.J.; Swain, C.J.; Ball, R.G.; *Bioorg. Med. Chem. Lett.*, **1994**, 4, 2545.
9. Natsugari, H.; Ikeura, Y.; Kiyota, Y.; Ishichi, Y.; Ishimaru, T.; Saga, O.; Shirafuji, H.; Tanaka, T.; Kamo, I.; Doi, T.; Otsuka, M.; *J. Med. Chem.*, **1995**, 38, 3106.
10. Elliott, J.; Seward, E.M.; *Exp. Opin. Ther. Patents*, **1997**, 7, 43.
11. Sisto, A.; Bonelli, F.; Centini, F.; Fincham, C.I.; Potier, E.; Monteagudo, E.; Lombardi, P.; Arcamone, F.; Goso, C.; Manzini, S.; Giolitti, A.; Maggi, C.A.; Venanzi, M.; Pispisa, B.; *Biopolymers*, **1995**, 36, 511.
12. Desai, M.C.; Vincent, L.A.; Rizzi, J.P.; *J. Med. Chem.*, **1994**, 37, 4263.
13. MacroModel V4.5 using Monte-Carlo searching and MM2 forcefield; Mohamadi, F.; Richards, N.G.J.; Guida, W.C.; Liskamp, R.; Lipton, M.; Caufield, C.; Chang, G.; Hendrickson, T.; Still, W.C.; *J. Comp. Chem.*, **1990**, 11, 440.
14. Hamor T.A.; Martin, I.L.; In *Progress in Medicinal Chemistry*; Ellis, G.P.; West G.B.; Eds.; Elsevier: New York, 1983; pp 193-205.
15. A similar observation was made regarding the importance of N-methylation in the related compounds of reference 9.

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